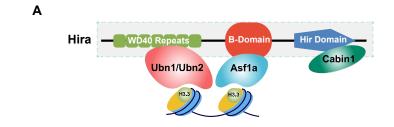
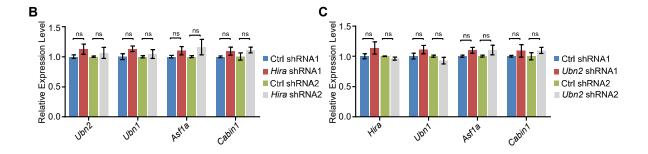
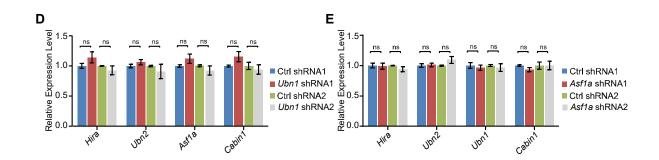
Supplementary Information

Supplementary Figure S1-8

Supplementary Table S1







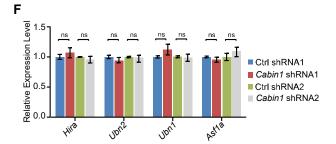


Fig. S1. The depletion of components of Hira complex does not disturb each other at mRNA level

- (A) Schematic diagram of the interrelation of HIRA members. Hira subunit associates with H3.3 through Ubn1/2 and Asf1a via its WD40 domain and B domain respectively.
- **(B)** qPCR analysis of the expression of *Ubn2*, *Ubn1*, *Asf1a*, and *Cabin1* after transfected with control shRNA and shRNA against *Hira*.
- (C) qPCR analysis of the expression of *Hira*, *Ubn1*, *Asf1a*, and *Cabin1* after transfected with control shRNA and shRNA against *Ubn2*.
- **(D)** qPCR analysis of the expression of *Hira*, *Ubn2*, *Asf1a*, and *Cabin1* after transfected with control shRNA and shRNA against *Ubn1*.
- (E) qPCR analysis of the expression of *Hira*, *Ubn2*, *Ubn1*, and *Cabin1* after transfected with control shRNA and shRNA against *Asf1a*.

(F) qPCR analysis of the expression of *Hira*, *Ubn2*, *Ubn1*, and *Asf1a* after transfected with control shRNA and shRNA against *Cabin1*. The results in B to F were normalized to *Gapdh*. Data are represented as mean \pm s.e.m. (n = 3 independent experiments) for the above qPCR results. ns: non-significant in Student's *t*-test.

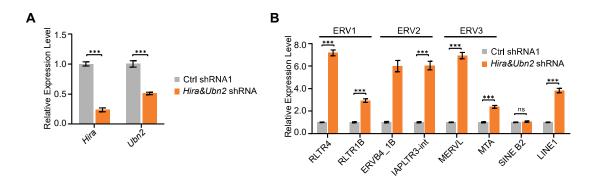


Fig. S2. Hira and Ubn2 double-knockdown in ESCs.

- (A) qPCR analysis of the expression of *Hira* and *Ubn2* of double-knockdown both *Hira* and *Ubn2* by shRNA in ESCs at the same time. ***p < 0.001 in Student's *t*-test.
- **(B)** qPCR analysis of the expression of different subfamilies endogenous retroviruses in *Hira & Ubn2* shRNA in ESCs. ns: non-significant, ***p < 0.001 in Student's *t*-test.

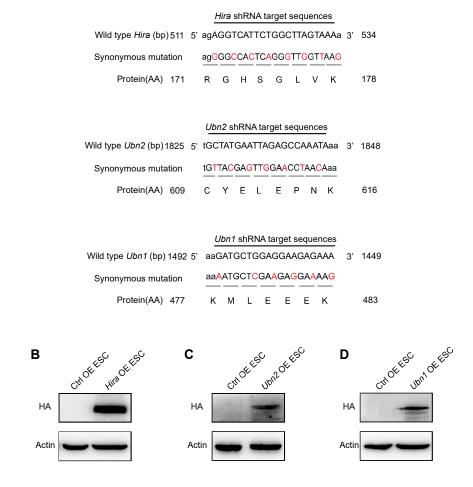


Fig. S3. Overexpression of shRNA-resistant *Hira*, *Ubn2*, and *Ubn1* in ESCs.

- (A) A schematic of shRNAs targeting sequences, synonymous mutation sequences and corresponding protein of *Hira*, *Ubn2* and *Ubn1*. Mutated nucleotide is highlighted in red; positions of nucleotide in gene are indicated on top.
- **(B-D)** Western blot analysis of Hira (B), Ubn2 (C), and Ubn1 (D) from overexpression ESCs with anti-HA antibody. Actin was included as a loading control.



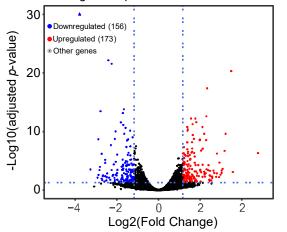


Fig. S4. Genome-wide expression changes after the knockdown of *Ubn1*.

The volcano plot of gene expression in Ubn1-depleted ESCs versus control ESCs. Significantly upregulated genes were labeled in red and significantly downregulated genes were labeled in blue. Horizontal blue dash line marked adjusted P-value (Wald test) 0.05 and vertical lines marked expression fold change 1.5. Triangles represent TEs with -log10 (adjusted P-value) > 30.

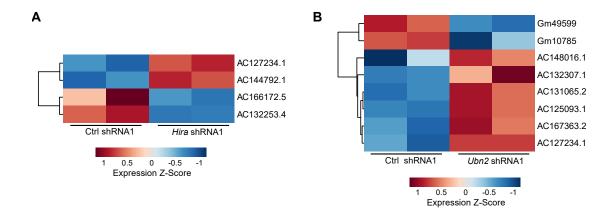


Fig. S5. Hira and Ubn2 regulate the expression of transposon-derived lncRNAs. (A-B) Heatmap of RNA-Seq expression Z-scores for transposon-derived lncRNAs that are differentially expressed in *Hira* (A) and *Ubn2* (B)-depleted ESCs versus control ESCs. Upregulated and downregulated genes are represented with red and blue colors respectively.

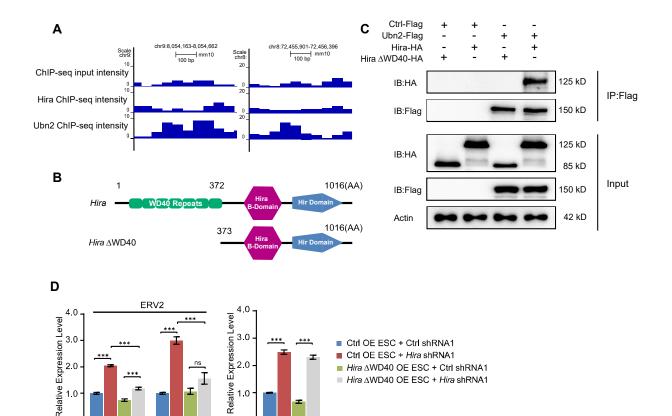


Fig. S6. Hira regulates MERVL through the interaction with Ubn2.

MERVI

0.0

ERVER 18

PLIP/2H

- (A) According to the published ChIP-seq data described in Methods, the enrichment of Hira and Ubn2 in the MT2/MERVL region is exemplified. Inputs are included as controls.
- **(B)** A schematic summary of *Hira* Δ WD40 mutant used for rescue. The length of the WD40 mutant form is indicated at the top in amino acids (AA). Δ , deletion.
- (C) Co-immunoprecipitation results confirmed that Ubn2 combined with Hira at its WD40 domain. The plasmids of Ctrl-Flag/*Ubn2*-Flag/*Hira*-HA/*Hira* ΔWD40-HA were respectively transfected into HEK 293T cells and analyzed via western blot of anti-Flag immunoprecipitation.
- **(D)** qPCR analysis of ERVB4_1B, RLTR12H, and MERVL in *Hira*-depleted ESCs after overexpression of *Hira* Δ WD40. qPCR results are normalized to *Gapdh*. Data are represented as mean \pm s.e.m. (n = 3 independent experiments). ns: non-significant, ***p < 0.001 in Student's *t*-test.

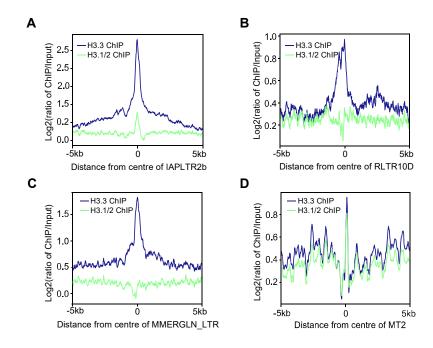


Fig. S7. H3.3 is enriched on three classes of ERVs. (A-D) H3.3 (blue) and H3.1/H3.2 (green) binding profile around the center of IAPLTR2b (A), RLTR10D (B), MMERGLN_LTR (C), and MT2 (D) locus in ESCs. The ChIP-seq signal was calculated as the log2 ratio of the normalized number of reads relative to the input.

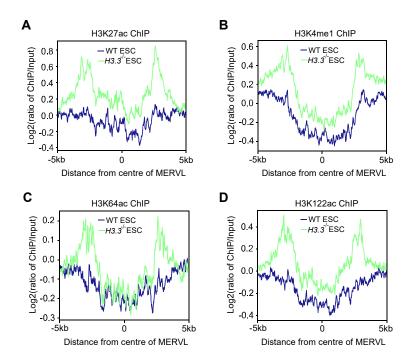


Fig. S8. Enrichment of histone marks on MERVL after *H3.3* **knockout. (A-D)** Several histone marks (H3K27ac (A), H3K4me1 (B), H3K64ac (C), H3K122ac (D)) binding profile around the center of MERVL locus in WT ESCs (blue) and *H3.3*^{-/-} ESCs (green). The ChIP-seq signal was calculated as the log2 ratio of the normalized number of reads relative to the input.

Table S1. Sequences of primers and shRNAs

Table 51. Sequences of primers and shirtings	
Gene	Sequence F
Gapdh	AGAAACCTGCCAAGTATGATGAC
Oct4	GTGGAAAGCAACTCAGAGG
Sox2	GCGGAGTGGAAACTTTTGTCC
Nanog	TTGCTTACAAGGGTCTGCTACT
Hira	TGGTCGGAGGAGAATCACG
Ubn1	CTATGCCTGAGCAGGTAGCC
Ubn2	CTGCCTCAGGGTCTTCAGTG
Asf1a	CACCGAATGCAGGACTCATC
Cabin1	TCGCCACTCAGACTTGGAAC
Cdx2	AGGCTGAGCCATGAGGAGTA
Eomes	CAATGTTTTCGTGGAAGTGG
Fgfr2	CCTCGATGTCGTTGAACGGTC
MERVL	AAGAGCCAAGACCTGCTGAG
MT2	CTCTACCACTTGGACCATATGAC
SINEB1	GTGGCGCACGCCTTTAATC
SINEB2	GAGTAAGAGCACCCGACTGC
LINE1	GGACCAGAAAAGAAATTCCTCCC
RLTR1B	GGTCCACACAAACACCTACCTT
MTA	TCTGTGGGATGTTGTGTAGGAG
IAPLTR2b	CACATTCGCCGTTACAAGAT
MMERGLN_LTR	GAGCTTTGAAACCTGGGGCT
RLTR12H	GCTGAACAGCCAATGACTGG
RLTR10D	GACTGCAGCCAAGTCTTATG
RLTR4	AGCGTTAATTTGGTCAAAGTCT
ERVB4_1B	ATGGAGATATTCTTAGCTCTG
IAPLTR3-int	GCGGTACAAGACTGGCTTAA
MT2 ChIP-qPCR	GGCTACACCTTCTGCTGGAG

shRNA Sequence Gene

Control shRNA GATGAAATGGGTAAGTACA Hira shRNA1 AGGTCATTCTGGCTTAGTAAA Hira shRNA2 CAGGACCGTTAGCCATAAT Ubn1 shRNA1 GATGCTGGAGGAAGAGAAA Ubn1 shRNA2 CGGAAGAAATTCCAGTGGAAT Ubn2 shRNA1 GCTATGAATTAGAGCCAAATA Ubn2 shRNA2 **GGTGCTACTAAACCGTTGT** GTGGGCTCTGCAGAAAGTGAA Asf1a shRNA1 Asf1a shRNA2 GGGTAACAGTTGTTCTGAT GACCACGATTACGTCAAAT Cabin1 shRNA1 GGAGGAGATAAGTCTAAGA Cabin1 shRNA2

Sequence R

GTCATTGAGAGCAATGCCAG GGTTCCACCTTCTCCAACT CGGGAAGCGTGTACTTATCCTT ACTGGTAGAAGAATCAGGGCT GAGGGTGACGATGCAGCAG GATCTTCACCACCTGGCACA CCCAGCATCCCAAAAGGAGT GCATCTGTTGAAAGAAGGGACTG TAGTGGGAGCAGCAGTTGTG **TGAGGTCCATAATTCCACTCA** GTTAGGAGATTCTGGGTGAA CAGCATCCATCTCCGTCACA TCCTCGTTTCTGCAACTGGT GAGGCTCCAAACAGCATCTCTA GACAGGGTTTCTCTGTGTAG AGAAGAGGGAGTCAGATCTCGT TTTGAGATACACCCTTCGAGGT CCACAGATCTTCACAATCCAAA TTGCTTACATCTTCAGGAGC AAACATCAGCAGCCTGTAAC CATGCCCGACCTCATGGCGA TCAGCCCAGTCCGCGTAACA CCAAGTATTGGGGACTGATAAT GAATTGACAGACATATGGAC GAACAGCTCCTCTTGCACGT TCGCAGCTGTGAATGGAAGT

Reference

Chen, et al. (2020) Nucleic acids res, 48, 10211-10225. Chen, et al. (2020) Nucleic acids res, 48, 10211-10225. Chen, et al. (2020) Nucleic acids res, 48, 10211-10225. Chen, et al. (2020) Nucleic acids res, 48, 10211-10225.

Zhang, et al. (2019) Nucleic acids res. 47, 8485-8501. Zhang, et al. (2019) Nucleic acids res. 47, 8485-8501. Chen, et al. (2020) Nucleic acids res, 48, 10211-10225. Chen, et al. (2021) Stem cells int, 2021, 6657597. CG CTCTTCTGGCTTTCATAGTCTCTGG Chen, et al. (2021) Stem cells int, 2021, 6657597. Zhang, et al. (2019) Nucleic acids res. 47, 8485-8501. Zhang, et al. (2019) Nucleic acids res, 47, 8485-8501.